



ez **accu shot**[™]
GROWTH PROMOTION TESTING

Verification Protocol Guide

MicroBioLogics[®]

MicroBioLogics®
Verification Protocol Guide

Accuracy & Traceability

have never been easier or more affordable.



Table of Contents

1. Introduction	3
1.1 Safety Guidelines	4
1.2 Available Technical Information	4
1.3 References	4
Acknowledgments	5
2. Verification Plan	5
2.1 Intended Location of Use	5
2.2 Microorganisms to be Tested	5
2.3 Scope of Verification	6
2.4 Verification Tests	6
2.5 Acceptance Criteria	7
2.6 Technician (s) Performing Verification	7
2.7 Approval of Plan	8
3. Verification of Traceability, Counts and Purity	8
3.1 Materials Needed for Verification	8
3.2 Medium Information	9
3.3. Documentation of Traceability to Reference Culture	9
3.4 Counts and Purity	10
3.5 Stability	11
3.6 Macroscopic and Microscopic Characteristics	13
3.7 Biochemical and/or Molecular Identity (If Required)	15
4. Summary and Conclusion	15
4.1 Summary of Data	15
4.2 Comments	16
4.3 Conclusion	17
4.4 Review and Approval	17

MicroBioLogics®

Verification Protocol Guide

1. Introduction

The purpose of this Verification Guide is to help customers smoothly and efficiently verify that EZ-Accu Shot™ microorganism strains are suitable for use in the Growth Promotion Test described in the United States Pharmacopeia (USP), the European Pharmacopoeia (EP), and the Japanese Pharmacopeias (JP). EZ-Accu Shot™ microorganisms are lyophilized, standardized, stable suspensions that deliver less than 100 CFU (colony forming units) when used as directed. Once verified, EZ-Accu Shot™ microorganism strains can be used as an easy and precise alternative to the seed-lot system and the time-consuming process of inoculum preparation.

EZ-Accu Shot™ microorganism strains meet the following pharmacopeial requirements:

1. Each strain delivers less than 100 CFU per 0.1 mL when used as directed.
2. Each strain is derived from a reference culture strain recommended by the harmonized pharmacopeias for growth promotion testing. The strains are traceable to the American Type Culture Collection (ATCC®). MicroBioLogics® is licensed by ATCC® to sell derivatives of ATCC® original microorganism strains.
3. Each strain is less than five passages from reference culture (the original master strain).
4. The identity of each strain has been confirmed by MicroBioLogics™.

In addition to meeting pharmacopeial requirements, EZ-Accu Shot™ microorganisms have the following benefits:

1. Lyophilized microorganism preparations dissolve in seconds.
2. There is little preparation time.
3. Microorganism suspensions can be used for eight hours if refrigerated after rehydration.
4. A Certificate of Assay is provided with every lot. The certificate lists the mean assay value (CFU concentration) obtained at MicroBioLogics®.
5. A Certificate of Analysis can be downloaded for every lot from the MicroBioLogics® website. The certificate lists such information as reference number, number of passages from reference culture, and macroscopic, microscopic and phenotypic characteristics when appropriate.

The Verification Guide has been divided into three parts.

Part 1: Verification Plan

- The laboratory determines the scope of the verification.
- The laboratory determines acceptance criteria.
- The plan is approved by management.

Part 2: The laboratory verifies the following for each EZ-Accu Shot™ strain:

- Traceability. Strains should be traceable to an original reference culture strain recommended by the pharmacopeias and should be less than five passages from the original master strain.
- Counts. Counts should be less than 100 CFU per agar plate, when product is used as directed.
- Purity. Contamination should not be seen on agar plates inoculated with the EZ-Accu Shot product.
- Stability. Microorganism suspensions should be stable for 8 hours after pellets are hydrated. Laboratories do not have to test stability if they do not intend to refrigerate microorganism suspensions.
- Identity. Identification methods can range from confirming microscopic and macroscopic microorganism characteristics to performing biochemical or molecular tests.

Part 3: Verification Summary

- The laboratory summarizes the data and determines if EZ-Accu Shot™ strains meets acceptance criteria.
- The laboratory submits the Verification Protocol to management for approval.

MicroBioLogics®

Verification Protocol Guide

1.1 Safety Guidelines

EZ-Accu Shot™ preparations contain potentially harmful microorganisms. All microorganisms produced by MicroBioLogics® are BioSafety Level 1 or 2. Guidelines for handling microorganisms can be found on the MicroBioLogics website in a Technical Bulletin called “Microorganism Biosafety Levels”. Directions for handling a microorganism spill can be found in the EZ-Accu Shot™ product insert.

Before using the EZ-Accu Shot™ product the MSDS should be downloaded and filed electronically or on site. There are two MSDS sheets available for the product. One MSDS is for Lyophilized Microorganism Preparations and the other MSDS is for Microorganism Hydrating Fluid.

1.2 Available Technical Information

The following documents are available to help with your validation.

- MSDS for Lyophilized Microorganism Preparations, MSDS.244
- MSDS for Microorganism Hydrating Fluid, MSDS.208
- “Microorganism Biosafety Levels”, TIB.072
- EZ- Accu Shot™ Product Insert, PI.276
- EZ-Accu Shot™ Illustrated Instructions, LIT. 280
- “Growth Promotion Test Guide for EZ-CFU™, EZ-CFU One Step™ and EZ-Accu Shot™”, TIB.264
- ATCC® 16404™ Name Change Letter, LIT.256 The Name Change Letter explains why the name of ATCC® 16404™ was changed to *Aspergillus brasiliensis*.

Documents are available at www.microbiologics.com. An easy way to obtain the documents is to use the search box. Search for documents by entering key words such as MSDS or biosafety.

1.3 References

The references used to create this guide are listed below.

1. The United States Pharmacopeia-National Formulary. 2009. <61> Microbiological Examination of Nonsterile Products: Microbial Enumeration Tests, First Supplement to USP32-NF27
2. The United States Pharmacopeia-National Formulary. 2009. <62> Microbiological Examination of Nonsterile Products: Tests for Specified Microorganisms, First Supplement to USP32-NF27
3. The United States Pharmacopeia-National Formulary. 2009. <71> Sterility Tests, First Supplement to USP32-NF27
4. The United States Pharmacopeia-National Formulary. 2009. <1226> Verification of Compendial Procedures FDA U.S. Food and Drug Administration. CFR - Code of Federal Regulations Title 21. 2009-04-01. Laboratory Records 21 CFR 211.194(a)(2)
5. The United States Pharmacopeia-National Formulary. 2009. <1111> Microbiological Examination of Nonsterile Products: Acceptance Criteria for Pharmaceutical Preparations and Substances for Pharmaceutical Use
6. European Pharmacopoeia. 2009. 2.6.12, Microbiological Examination of Nonsterile Products: Microbial Enumeration Tests.

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Verification Protocol Guide

7. European Pharmacopoeia. 2009. 2.6.13, Microbiological Examination of Nonsterile Products: Tests for Specified Microorganisms.
8. European Pharmacopoeia. 2009. 2.6.1, Sterility Tests.
9. Japanese Pharmacopoeia. May.19. 2009. 4.05, Microbiological Examination of Nonsterile Products: Microbial Enumeration Tests and Microbiological Examination of Nonsterile Products: Tests for Specified Microorganisms. 15th Partial Revision (English version)
10. Japanese Pharmacopoeia. May. 19. 2009. 4.06, Sterility Tests. 15th Partial Revision (English version)

Acknowledgements



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2. Verification Plan

2.1 Intended Location of Use

Company Name: _____

Address: _____

Department: _____

Signature: _____ **Date:** _____

2.2 Microorganisms to be Tested

In Table 1, check (✓) EZ-AccuShot™ microorganisms included in the verification study.

Table 1: EZ-Accu Shot™ Microorganisms

Microorganism	ATCC® number	EZ-Accu Shot™ Product number	Check (✓)if included in study
<i>Aspergillus brasiliensis</i>	ATCC® 16404™*	0392A	
<i>Bacillus subtilis</i> subsp. <i>spizizenii</i>	ATCC® 6633™*	0486A	
<i>Candida albicans</i>	ATCC® 10231™*	0443A	
<i>Clostridium sporogenes</i>	ATCC® 19404™*	0317A	
<i>Clostridium sporogenes</i>	ATCC® 11437™*	0487A	
<i>Escherichia coli</i>	ATCC® 8739™*	0483A	
<i>Pseudomonas aeruginosa</i>	ATCC® 9027™*	0484A	
<i>Staphylococcus aureus</i> subsp. <i>aureus</i>	ATCC® 6538™*	0485A	

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Verification Protocol Guide

2.3 Scope of Verification

Briefly summarize the purpose of the Verification.

2.4 Verification Tests

List the Tests which the laboratory will perform in order to verify the EZ-Accu Shot™ product.

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Verification Protocol Guide

2.5 Acceptance Criteria

Describe acceptance criteria for determining suitability of EZ-Accu Shot™ microorganisms.

2.6 Technician(s) Performing Verification

Name and signature of technician(s) performing validation.

Name (print) _____ Title _____

Signature _____ Date _____

Department _____

Company Name _____

Address _____

Name (print) _____ Title _____

Signature _____ Date _____

Department _____

Company Name _____

Address _____

Name (print) _____ Title _____

Signature _____ Date _____

Department _____

Company Name _____

Address _____

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Verification Protocol Guide

Name (print) _____ Title _____
Signature _____ Date _____
Department _____
Company Name _____
Address _____

2.7 Approval of Plan

Signature(s) of management staff

Name (print) _____ Title _____
Signature _____ Date _____

Name (print) _____ Title _____
Signature _____ Date _____

Name (print) _____ Title _____
Signature _____ Date _____

Name (print) _____ Title _____
Signature _____ Date _____

3. Verification of Traceability, Counts, and Purity

3.1 Materials Needed for Verification

3.1.1 The following materials are needed for testing counts and purity.

- EZ-Accu Shot™ lyophilized microorganism preparations
- Hydrating Fluid (as received with each EZ-Accu Shot™ kit)
- Calibrated pipetter and sterile tips
- Media such as Soybean Casein Agar (Tryptic Soy Agar) and Sabouraud Dextrose Agar
- Vortex Mixer
- Sterile Spreaders
- Incubator at 20-25°C
- Incubator at 30-35°C
- Glass slides
- Gram Stain Kit for staining bacteria
- Lactophenol Blue Stain and clear tape for staining *Aspergillus brasiliensis*

3.1.2 Macroscopic characteristics should match the description of the microorganisms provided in Table 10. If your protocol requires further identification testing, you will need a manual or automated biochemical or molecular system.

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Verification Protocol Guide

3.2 Medium Information

In Table 2, record the lot number and expiration date of the agar used for growing the microorganisms. Certificates of Analysis for each medium should be attached to the Verification paper work. Check (✓) if Certificate of Analysis is attached.

Table 2: Media used for Verification

Medium	Manufacturer	Catalog Number	Lot Number	Expiry Date	Certificate of Analysis (✓)	Recorded By
Sabouraud Dextrose Agar						
Soybean Casein Agar (Tryptic Soy Agar)						
Columbia Blood Agar						

3.3 Documentation of Traceability to Reference Culture

In Table 3, record the lot number and the expiration date of each microorganism strain tested. Record the Hydrating Fluid lot number and the expiry date. Check the appropriate column after the Certificates of Assay and Analysis have been attached to the Verification paperwork. Both certificates document traceability to the original reference culture and the passage number from reference culture.

Table 3: Documentation of Microorganism and Hydrating Fluid Lot Numbers

Microorganism	Catalog No.	Microorganism Lot Number	Expiry Date	Hydrating Fluid Lot Number	Expiry Date	Certificate of Assay (✓)	Certificate of Analysis (✓)	Recorded by
<i>Aspergillus brasiliensis</i>	0392A							
<i>Bacillus subtilis</i> subsp. <i>spizizenii</i>	0486A							
<i>Candida albicans</i>	0443A							
<i>Clostridium sporogenes</i>	0317A							
<i>Clostridium sporogenes</i>	0487A							
<i>Escherichia coli</i>	0483A							
<i>Pseudomonas aeruginosa</i>	0484A							
<i>Staphylococcus aureus</i> subsp. <i>aureus</i>	0485A							

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Verification Protocol Guide

3.4 Counts and Purity

For each EZ-Accu Shot™ strain, do the following:

1. Remove one vial of Hydrating Fluid and one foil pouch from the canister. Remove vial containing the lyophilized microorganism pellet from the foil pouch. Allow unopened vials to equilibrate to room temperature.
2. Label Hydrating Fluid vial with microorganism lot number.
3. Remove screw top and stopper from Hydrating Fluid.
4. Remove screw top from vial containing lyophilized pellet.
5. Tip pellet into Hydrating Fluid.
6. Vortex 5 seconds or until suspension is homogenous.
7. Dispense 0.1 mL (100 µl) of suspension per agar plate. Perform a total of five replicates.
8. Spread inoculum evenly across plate with sterile spreader.
9. Incubate agar plates. Follow directions in Table 4 for temperature and length of time for incubation. Record incubator identity, incubator temperature range, date and time when incubation begins and initials in Table 5.
10. Recap the microorganism vial and store the suspension at 2-8°C if intending to verify 8 hour stability. (This step is not necessary if the laboratory never plans to store the suspension.) Record refrigerator identity, refrigerator temperature range, date and time suspension is placed in the refrigerator, and initials in Table 7 in Section 3.5, Verification of Stability.
11. Record in Table 5 the date and time when incubation ends. Record initials.
12. Count the colonies on each replicate plate and record the number in Table 6. Determine and record average count.
13. Inspect the plates for purity. If no contamination is seen, record S in the Purity Column in Table 6. If contamination is seen, record U. Date and initial.

Table 4: Incubation Instructions

EZ-Accu Shot™ Microorganism	Product Number	Media	Temperature Range	Pharmacopeia Incubation Instructions	Check Macroscopic Characteristics	Atmosphere
<i>Aspergillus brasiliensis</i>	0392A	Soybean Casein Agar ¹	30-35°C	≤5 days	4-5 days	Aerobic
<i>Aspergillus brasiliensis</i>	0392A	Sabouraud Dextrose Agar	20-25°C	≤5 days	4-5 days	Aerobic
<i>Bacillus subtilis</i> subsp. <i>spizizenii</i>	0486A	Soybean Casein Agar ¹	30-35°C	≤3 days	24 hours	Aerobic
<i>Candida albicans</i>	0443A	Soybean Casein Agar ¹	30-35°C	≤5 days	48 hours	Aerobic
<i>Candida albicans</i>	0443A	Sabouraud Dextrose Agar	20-25°C	≤5 days	48 hours	Aerobic
<i>Clostridium sporogenes</i>	0317A	Columbia Blood Agar	30-35°C	48 hours	48 hours	Anaerobic
<i>Clostridium sporogenes</i>	0487A	Columbia Blood Agar	30-35°C	48 hours	48 hours	Anaerobic
<i>Escherichia coli</i>	0483A	Soybean Casein Agar ¹	30-35°C	≤3 days	24 hours	Aerobic
<i>Pseudomonas aeruginosa</i>	0484A	Soybean Casein Agar ¹	30-35°C	≤3 days	24 hours	Aerobic
<i>Staphylococcus aureus</i> subsp. <i>aureus</i>	0485A	Soybean Casein Agar ¹	30-35°C	≤3 days	24 hours	Aerobic

¹ Soybean Casein Agar is also known as Tryptic Soy Agar

MicroBioLogics®

Verification Protocol Guide

Table 5: Incubation Data

Microorganism Name	Catalog Number	Incubator Identity	Incubator Temperature Range	Date and Time Incubation Begins	By	Date and Time Incubation ends	By
<i>Aspergillus brasiliensis</i>	0392A						
<i>Bacillus subtilis</i> subsp. <i>spizizenii</i>	0486A						
<i>Candida albicans</i>	0443A						
<i>Clostridium sporogenes</i>	0317A						
<i>Clostridium sporogenes</i>	0487A						
<i>Escherichia coli</i>	0483A						
<i>Pseudomonas aeruginosa</i>	0484A						
<i>Staphylococcus aureus</i> subsp. <i>aureus</i>	0485A						

Table 6: Microorganism Counts and Purity

Microorganism	Catalog Number	Type of Agar ¹	CFU Count					Avg. Count	Purity S/U	Date	By
			1	2	3	4	5				
<i>A. brasiliensis</i>	0392A	SCA									
<i>A. brasiliensis</i>	0392A	SDA									
<i>B. subtilis</i> subsp. <i>spizizenii</i>	0486A	SCA									
<i>C. albicans</i>	0443A	SCA									
<i>C. albicans</i>	0443A	SDA									
<i>C. sporogenes</i>	0317A	CBA									
<i>C. sporogenes</i>	0487A	CBA									
<i>E. coli</i>	0483A	SCA									
<i>P. aeruginosa</i>	0484A	SCA									
<i>S. aureus</i> subsp. <i>aureus</i>	0485A	SCA									

¹SCA = Soybean Casein Agar, SDA = Sabouraud Dextrose Agar, CBA = Columbia Blood Agar

3.5 Stability

For each EZ-Accu Shot™ strain, do the following:

1. Remove suspension from refrigerator after 8 hours (or less if laboratory is planning for shorter storage duration). Record date and time when the suspension is taken out of the refrigerator and record initials in Table 7.

MicroBioLogics®
Verification Protocol Guide

2. Remix suspension on vortex mixer.
3. Dispense 0.1 mL of suspension per plate. Set up a total of five replicates.
4. Spread inoculum evenly across plate with sterile spreader.
5. Incubate second set of agar plates at the same temperature and for the same length of time as the first set of agar plates. Record incubator identity, incubator temperature range, date and time incubation begins, and initials in Table 8.
6. Record the date and time when incubation ends and initials in Table 8.
7. Count the colonies on each replicate plate. Record the number in Table 9. Determine and record average count. Record date and initials.

Table 7: Refrigeration Data

Microorganism Lot Number	Catalog Number	Refrigerator Identity	Refrigerator Temperature Range	Date and Time Refrigeration Begins	By	Date and Time Refrigeration Ends	By
<i>Aspergillus brasiliensis</i>	0392A						
<i>Bacillus subtilis</i> subsp. <i>spizizenii</i>	0486A						
<i>Candida albicans</i>	0443A						
<i>Clostridium sporogenes</i>	0317A						
<i>Clostridium sporogenes</i>	0487A						
<i>Escherichia coli</i>	0483A						
<i>Pseudomonas aeruginosa</i>	0484A						
<i>Staphylococcus aureus</i> subsp. <i>aureus</i>	0485A						

Table 8: Incubation Data for Refrigerated Suspensions

Microorganism Lot Number	Catalog Number	Incubator Identity	Incubator Temperature Range	Date and Time Incubation Begins	By	Date and Time Incubation Ends	By
<i>Aspergillus brasiliensis</i>	0392A						
<i>Bacillus subtilis</i> subsp. <i>spizizenii</i>	0486A						
<i>Candida albicans</i>	0443A						
<i>Clostridium sporogenes</i>	0317A						
<i>Clostridium sporogenes</i>	0487A						
<i>Escherichia coli</i>	0483A						
<i>Pseudomonas aeruginosa</i>	0484A						
<i>Staphylococcus aureus</i> subsp. <i>aureus</i>	0485A						

MicroBioLogics®
Verification Protocol Guide

Table 9: Microorganism counts after refrigeration

Microorganism	Catalog Number	Type of Agar	CFU Count					Avg. Count	Date	By
			1	2	3	4	5			
<i>Aspergillus brasiliensis</i>	0392A	SCA								
<i>Aspergillus brasiliensis</i>	0392A	SDA								
<i>Bacillus subtilis</i> subsp. <i>spizizenii</i>	0486A	SCA								
<i>Candida albicans</i>	0443A	SCA								
<i>Candida albicans</i>	0443A	SDA								
<i>Clostridium sporogenes</i>	0317A	CBA								
<i>Clostridium sporogenes</i>	0487A	CBA								
<i>Escherichia coli</i>	0483A	SCA								
<i>Pseudomonas aeruginosa</i>	0484A	SCA								
<i>Staphylococcus aureus</i> subsp. <i>aureus</i>	0485A	SCA								

¹SCA = Soybean Casein Agar, SDA = Sabouraud Dextrose Agar, CBA = Columbia Blood Agar

3.6 Macroscopic and Microscopic Characteristics

1. After incubation examine colonies from all strains macroscopically. Colony appearance should agree with descriptions listed in Table 10. Enter S (satisfactory appearance) or U (unsatisfactory appearance) in appropriate column. Record date and initials.
2. Use the Gram Stain method to stain bacterial colonies from each strain. Examine all strains microscopically. Microscopic appearance should agree with descriptions listed in Table 10. Enter S (satisfactory appearance) or U (unsatisfactory appearance) in appropriate column. Record date and initials.
3. Examine *Aspergillus brasiliensis* microscopically after spores have formed (this may take 5-7 days). To do this, firmly press the sticky side of transparent tape to the surface of the fungal culture. Gently place the tape on a drop of Lactophenol Blue on a clear glass slide. Examine slide microscopically (a magnification of 400X works well). Microscopic appearance should agree with descriptions listed in Table 10. Enter S (satisfactory appearance) or U (unsatisfactory appearance) in appropriate column. Record date and initials.
4. If SOP calls for further testing, subculture colonies from each strain onto fresh agar plates (the new plates are the fifth passage from Reference Culture).
5. After incubation, perform identification tests, if required to do so by SOP. Describe identification method in Section 3.7.
6. Record identification in Table 11. Enter S (satisfactory results) or U (unsatisfactory results) in appropriate column. Record date and initials. Attach worksheets and results.
7. Autoclave contaminated articles or dispose of them in a biohazard container.

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Verification Protocol Guide

Table 10: Macroscopic and Microscopic Characteristics

Strain	Cat. #	Agar ¹	Macroscopic description	S/U	Date	By	Microscopic description	S/U	Date	By
<i>A. brasiliensis</i>	0392A	SCA, 30-35°C, 4-5 days.	By 4 days, rapidly growing colonies which are initially white or pale yellow, quickly become black with conidia (spore) production. Reverse is pale yellow.				Lactophenol Blue Stain: chains of small conidia which arise from short sterigmata arranged radially over the surface of the vesicle.			
<i>A. brasiliensis</i>	0392A	SDA, 20-25°C, 4-5 days.	By 4 days, rapidly growing white colonies which quickly become black with conidia (spore) production.				Lactophenol Blue Stain: chains of small conidia which arise from short sterigmata arranged radially over the surface of the vesicle.			
<i>B.subtilis</i> subsp. <i>spizizenii</i>	0486A	SCA, 30-35°C, 24 hours.	Large, irregular, flat, undulate edge, cream and wrinkled with ground glass appearance; slight yellow coloring may appear in wrinkles by 48 hours.				Straight, gram positive rod, with an ellipsoidal, central, or terminal endospore.			
<i>C. albicans</i>	0443A	SCA, 30-35°C, 48 hours.	Large, white, circular, convex, dull colonies.				Gram positive, spherical, budding yeast cells.			
<i>C. albicans</i>	0443A	SDA, 20-25°C, 48 hours.	Large, white, circular, convex, dull colonies.				Gram positive, ovoid, budding yeast cells.			
<i>C. sporogenes</i>	0317A	CBA, anaerobic conditions, 30-35°C, 48 hours.	Large, irregularly circular, with coarse rhizoid edge, slightly raised center, gray, beta hemolytic, swarms, adheres to agar.				Gram positive straight rods. Spores, when seen, are oval, central or subterminal, and distend the cell slightly.			
<i>C. sporogenes</i>	0487A	CBA, anaerobic conditions, 30-35°C, 48 hours.	Large, irregularly circular, with coarse rhizoid edge, slightly raised center, gray, beta hemolytic, swarms, adheres to agar.				Gram positive straight rods. Spores, when seen, are oval, central or subterminal, and distend the cell slightly.			
<i>E. coli</i>	0483A	SCA, 30-35°C, 24 hours.	Medium to large, creamy, low convex, slightly rough, slightly erose.				Gram negative straight rod.			
<i>P. aeruginosa</i>	0484A	SCA, 30-35°C, 24 hours.	Medium to large, flat, circular to irregular shaped, cream.				Straight or slightly curved gram negative rod.			
<i>S. aureus</i> subsp. <i>aureus</i>	0485A	SCA, 30-35°C, 24 hours.	Medium, convex, circular, glistening, smooth, creamy, opaque, light gold colonies.				Gram positive cocci occurring singly, in pairs, and in irregular clusters.			

¹SCA = Soybean Casein Agar, SDA = Sabouraud Dextrose Agar, CBA = Columbia Blood Agar

MicroBioLogics®
Verification Protocol Guide

3.7 Biochemical and/or Molecular Identity (If Required)

Table 11: Identification Test Results

Microorganism	Catalog Number	Biochemical/Molecular Identification	S/U	Date	By
<i>Aspergillus brasiliensis</i>	0392A				
<i>Bacillus subtilis</i> subsp. <i>spizizenii</i>	0486A				
<i>Candida albicans</i>	0443A				
<i>Clostridium sporogenes</i>	0317A				
<i>Clostridium sporogenes</i>	0487A				
<i>Escherichia coli</i>	0483A				
<i>Pseudomonas aeruginosa</i>	0484A				
<i>Staphylococcus aureus</i> subsp. <i>aureus</i>	0485A				

4. Summary and Conclusion

Determine if each EZ-Accu Shot™ lyophilized microorganism strain meets acceptance criteria.

4.1 Summary of Data

Enter S (Satisfactory) or U (Unsatisfactory) for each category. Date and initial.

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Verification Protocol Guide

Table 12: Summary

Microorganism	Cat. no.	Type of media ¹	Traceability S/U	CFU count S/U	8 hour Stability S/U	Purity S/U	Identity S/U	Date	By
<i>Aspergillus brasiliensis</i>	0392A	SCA							
<i>Aspergillus brasiliensis</i>	0392A	SDA							
<i>Bacillus subtilis</i> subsp. <i>spizizenii</i>	0486A	SCA							
<i>Candida albicans</i>	0443A	SCA							
<i>Candida albicans</i>	0443A	SDA							
<i>Clostridium sporogenes</i>	0317A	CBA							
<i>Clostridium sporogenes</i>	0487A	CBA							
<i>Escherichia coli</i>	0483A	SCA							
<i>Pseudomonas aeruginosa</i>	0484A	SCA							
<i>Staphylococcus aureus</i> subsp. <i>aureus</i>	0485A	SCA							

¹SCA = Soybean Casein Agar, SDA = Sabouraud Dextrose Agar, CBA = Columbia Blood Agar

4.2 Comments

By _____ Date _____

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Verification Protocol Guide

4.3 Conclusion

By _____ Date _____

4.4 Review and Approval

Name	Title	Signature	Date