

# 96-Well G-50 Plate

*For research use only*

<b>Storage</b>	: 2-8°C for up to 6 months (Do not freeze)
<b>Volume</b>	: 20 to 50 µl
<b>Format</b>	: 96-well plates

**Geneaid**



[www.geneaid.com](http://www.geneaid.com)

## Introduction

96-Well G-50 Plates consist of pre-packed Sephadax G-50, pre-hydrated with double-distilled water. The plates are ideal for removing excess dye terminator, freeing nucleotides from sequencing and labeling reactions, desalting and buffer exchange. G-50 Plates can purify DNA fragments larger than 20 bases in length with low molecular weight material retained in the gel matrix. Since G-50 Plates are designed to purify DNA fragments >20 bases only, they are not recommended for PCR product primer removal.

### Kit Contents

Name	CGP02	CGP04	CGP10
G-50 Plate	2 pcs	4 pcs	10 pcs

### Order Information

Product Name	Package Size	Cat. No.
RNA Pure Kit	100/300 preps	PR100/300
DNA Pure Kit	100/300 preps	DP100/300
G-25 Mini Column	50 preps	CG025
G-50 Mini Column	50 preps	CG050
96-Well G-50 Plate	4/10 x 96 Wells	CGP04/10

## Specifications

In the event of gel drying (cracking), add 50-100 µl of ddH<sub>2</sub>O to each well of the 96-Well G-50 Plate before use. The optimal sample volume is 20 to 50 µl (50 µl maximum).

## Purification/Desalting Protocol

Step 1	<ul style="list-style-type: none"><li>Remove the adhesive film from the <b>G-50 Plate</b>.</li><li>Place the <b>G-50 Plate</b> on a 2 ml collection plate.</li><li>Centrifuge at 2,500 x g for 5 minutes.</li></ul>
Step 2	<ul style="list-style-type: none"><li>Transfer the <b>G-50 Plate</b> to a 0.35 ml collection plate.</li><li>Carefully load the sample (20-50 µl) onto the center of each gel bed surface.</li></ul>
Step 3	<ul style="list-style-type: none"><li>Centrifuge again at 2,500 x g for 5 minutes.</li><li>Each purified sample can be recovered at the bottom of the 0.35 ml collection plate (approximately the same volume as the loaded sample).</li></ul>

## Buffer Exchange Protocol

Step 1	<ul style="list-style-type: none"><li>Remove the adhesive film from the <b>G-50 Plate</b>.</li><li>Place the <b>G-50 Plate</b> on a 2 ml collection plate and centrifuge at 2,500 x g for 5 minutes.</li></ul>
Step 2	<ul style="list-style-type: none"><li>Discard the flow-through in the 2 ml collection plate and place the <b>G-50 Plate</b> back on the same 2 ml collection plate.</li></ul>
Step 3	<ul style="list-style-type: none"><li>Add 350 µl of desired buffer to each well of the <b>G-50 Plate</b>.</li><li>Centrifuge at 2,500 x g for 5 minutes.</li></ul>
Step 4	<ul style="list-style-type: none"><li>Transfer the <b>G-50 Plate</b> to a 0.35 ml collection plate.</li><li>Carefully load the sample (20-50 µl) onto the center of each gel bed surface.</li></ul>
Step 5	<ul style="list-style-type: none"><li>Centrifuge again at 2,500 x g for 5 minutes. The purified sample can be recovered at the bottom of the 0.35 ml collection plate (approximately the same volume as the loaded sample).</li></ul>

## Troubleshooting

Problem	Possible Reasons/Solution
Gel Drying	<ul style="list-style-type: none"><li>Add 50-100 µl of ddH<sub>2</sub>O to each well of the G-50 Plate before use.</li></ul>